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3. METHOD TO VISUALIZE LATENTLY-INFECTED T CELLS

• Generate a full-length HIV latency reporter to follow latently-infected T cells in realtime.



AIM

To investigate the effect of dendritic cells and IL-7 on latently infected CD4⁺T cell populations.

HYPOTHESIS

 IL-7 and signalling thorough DC:T cell interactions contribute to the maintenance of HIV latently infected CD4⁺T cells *in vivo*

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Figure 1:HIV Nef-CRMZY delineates between productive and latent HIV infections in CD4⁺ T cells

Naïve CD4+ T cells were isolated from PBMC and infected with HIV _{Nef-CRMZY} at an MOI of 0.1. 5days post infection, productively infected cells (red, E2Crimson⁺ZsGreen1⁺) and latently infected cells (green,E2Crimson⁻ZsGreen1⁺) were visualized by flow cytometry (A). CD4 downregulation in productively infected cells (red, E2Crimson⁺ZsGreen1⁺) and latently infected cells (green,E2Crimson⁻ZsGreen1⁺) was analyzed (B). latently infected cells (green,E2Crimson⁻ZsGreen1⁺) was analyzed for their CD4 expression (C). CD4⁺ latent T cells and CD4⁻ Latent T cells were analyzed for HIV gag (p24) (D). Results are representative of 5 independent experiments.

CD4⁺T cells were infected with HIV _{Nef-CRMZY} for 5days. After 5days, immunocult (CD3/CD28) cell activator was used to reactivate latently infected cells for 24hrs, activation status was confirmed by assessing CD69⁺HLA-DR⁺) (A). Activated latent cells were assessed for evidence of productive infection by evaluating p24 staining (B). Relative ratio of productive infected cells and latently infected cells were analysed (C).****, p < 0.001.

Figure 2: Latent HIV Nef-CRMZY infected CD4+ T cells can be reactivated

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p24⁻ZsGreen1⁺ latently infected T cells

Figure 3: Addition of autologous Dendritic cells and IL-7 synergize to expand the proportion of CD4⁺ p24⁻ ZsGreen1⁺ latently infected T cells

CD4⁺T cells were infected with HIV_{Nef-CRMZY} for 5 days. At 5 days post-infection, IL-7 (50ng/ml) and DCs (1 DC: 3 T cells) were added to the culture. 10 days later, the proportion of latently infected cells were assessed by flow cytometry (A). Percentage of CD4 expressing latently infected T cells were graphically represented (B). Relative ratio of latently infected (CD4+ and CD4- T cells) were assessed (C). ***, p < 0.001. ****, p < 0.0001.

CD4⁺T cells were infected with HIV_{Nef-CRMZY} for 5 days. At 5 days post-infection, IL-7 (50ng/ml) and DCs (1 DC: 3 T cells) were added to the culture. 10 days later, the population of proliferated (TagIT^{neg}) latently infected cells were assessed by flow cytometry (A). Population of latently infected T cells expressing pro-survival molecule (BCL-2) were assessed by flow cytometry (B).

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SIGNIFICANCE AND CONCLUSIONS	RELEVANCE OF OUR FINDING TO HIV STUDY
 We have developed a dual reporter HIV that encodes for all HIV proteins and allows us to follow latently infected T cells for up to 15 days in culture. Importantly, the expression of full-length Nef allows us to use CD4 downregulation as a sensitive marker of Nef expression and define latently-infected cells using our system as ZsGreen1⁺CD4^{high}p24^{neg}. We show that physiological DC:T cell interactions in the lymph node, along with IL-7 signaling, that help promote homeostatic proliferation of central memory T cells, may also be co-opted by latently infected T cells for their own survival and proliferation. This may help explain why latent T cells can persist for many years and represents a major barrier to achieve HIV cure in infected individuals unloss these mechanisms are disrupted. 	 Our data suggest that dendritic cells and IL-7 may synergize to facilitate latently-infected T cell expansion by promoting cell survival and/or proliferation. Our <i>in vitro</i> model system will help define cellular and molecular mechanisms that may lead to the expansion of the viral reservoir, with the possibility of testing whether antigen recognition also play a role in this process.

Dendritic cells and IL-7 synergize to increase latent-infected CD4⁺ T cell populations **Nnamdi Ikeogu**¹, Oluwaseun Ajibola¹, Xinyun Liu¹, Paul Lopez¹, Roshan Parvarchian¹, Alon Hershhorn² and Thomas Murooka¹

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FUTURE DIRECTIONS

- Evaluate the mechanism of DC mediated maintenance of CD4⁺ latent T cells (MHC-II blockade, CD80 and CD86 blockade)
- Visualize latently infected CD4⁺ T cells in the lymph node of humanized mice.
- Assess whether, antigens (e.g. flu, gut bacteria) preferentially expands latent T cells in the presence of dendritic cells and IL-7 without causing reactivation of latent T cells

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